Dr.ssa Giulia Genoni

Clinica Pediatrica, Università degli studi del Piemonte Orientale "A.Avogadro" Dottorato in Scienze e Biotecnologie Mediche (Percorso d'eccellenza)

Paediatric obesity and cardiovascular dysfunction: searching for early markers of damage

Background

During the past two decades, the prevalence of obesity in children has risen worldwide. Childhood obesity causes a wide range of serious complications, increasing the risk of premature morbidity and mortality and raising public-health concerns (1).

In addition, obese children are more prone to become obese adults, with an enhanced risk for cardiovascular diseases (CVD) (2,3). A cluster of cardiovascular disease risk factors as hypertension, dyslipidaemia, chronic inflammation, increased blood clotting tendency, endothelial dysfunction, and hyperinsulinaemia has been identified in children as young as 5 years of age (4). Among adolescents and young adults, the presence of cardiovascular disease risk factors correlates with asymptomatic coronary atherosclerosis, and lesions were more advanced in obese subjects (5-7). Moreover, recent studies reported that paediatric obesity is often associated with current cardiovascular abnormalities, suggesting that this issue requires immediate attention to prevent progressive cardiovascular damage.

Childhood obesity has been related to an impaired cardiac structure and function. Obese children show left atrial (LA) and left ventricular (LV) dimensions and mass significantly greater than lean controls (8-10). Moreover, greater epicardial fat has also been reported in children with obesity. Moreover, obese children and adolescents show an altered cardiac mechanics including diastolic dysfunction and systolic dysfunction at rest and during exercise (8-10, 11,12).

It is well known that atherogenesis and arterial wall damage begin during childhood. Furthermore, there is evolving evidence that clinical indicators of atherosclerosis such as CIMT, arterial stiffness, and endothelial dysfunction are altered in children with obesity, although mechanisms by which these effects are mediated have not been fully understood (13-17).

Therefore, the detection of a high cardiovascular risk profile associated to obesity during childhood would allow the early diagnosis of cardiovascular complications and the institution of preventive and therapeutic measures.

The mechanisms relating adiposity to cardiovascular dysfunction in children are difficult to ascertain (18). Obesity is a state of chronic low-level inflammation and increased oxidative stress.

Oxidative stress plays an important role in the pathogenesis of cardiovascular alterations by either triggering or exacerbating the biochemical processes accompanying endothelial dysfunction (19).

Moreover, adipose tissue acts as a secretory gland, releasing hormones and adipokines with pro- or anti-inflammatory activity (20). Clinical studies of obese adult subjects have observed an association between plasma levels of adipokines and markers of inflammation and/or oxidative stress (19).

Among various adipokines, adiponectin, the most abundant protein secreted by the adipose tissue and circulating in human plasma as multimeric forms, seems to play an important role for several reasons. First, in contrast to other adipokines, which are up-regulated in obesity, secretion of adiponectin is markedly reduced in obese subjects and in most experimental models of obesity. Second, adiponectin seems to exert mainly positive activities on metabolism, vascular tone and inflammatory reaction. Consequently, in contrast to other adipokines, which circulate in excess in obese subjects and exert unbeneficial effects when chronically elevated, deficiency rather than excess of adiponectin is implicated in obesity-associated complications. Finally, serum concentration of adiponectin is very high in comparison to other hormones and cytokines, which suggests that apart from binding to specific high-affinity receptors, this protein may also have some less specific low affinity targets (21).

Adiponectin has been associated with endothelial improvement and vascular protection through the activation of an endothelial isoform of nitric oxide (eNOS)–related signalling and with antiinflammatory properties and antiatherogenic effects (22,23). Moreover, adiponectin plays a protective role in myocardial ischemia-reperfusion injury and myocardial hypertrophy in human studies (21) and increase systolic and diastolic function and coronary blood flow in animal models (24). Conversely, adiponectin deficiency has been reported to be predictive of future adverse cardiac events and has been associated with increased oxidative stress and inferior cardiac function recovery (21).

Thus, an impaired production of adipokines, such as in obese children, may be a key mechanism linking obesity with inflammation and oxidative stress. Moreover, adiponectin may be one of the primary signals linking inflammation, obesity and cardiovascular dysfunction.

The understanding of these complex mechanisms and the identification of possible early markers of cardiovascular damage are therefore necessary in order to establish preventive and therapeutic measures in childhood and to decrease cardiovascular morbidity and mortality in adulthood.

Aims

In the present research study we aim to examine cardiac and vascular changes in obese children and adolescents and to analyze the relationship with plasmatic adipokines, inflammation and oxidative stress markers at baseline and after 6 and 12 months of an isocaloric Mediterranen balanced diet plus an exercise training regimen.

The first main endpoint will be the difference of 10% of diastolic function between controls and obese subjects.

The second endpoint will be the variation of 20% of diastolic function in the obese group after 6 months of treatment.

The third endpoint will be the evaluation of the association between cardiovascular dysfunction and adiponectin and, inflammatory and oxidative stress plasmatic markers.

Subjects

This study is a single-centre longitudinal study. Subjects will be recruited at Division of Pediatrics, Department of Health Sciences, University of Piemonte Orientale, Novara. All subjects will be evaluated at baseline (T0) and after 6 (T1) and 12 months (T2) of an isocaloric Mediterranen balanced diet plus aerobic training. The study protocol will be in accordance with the ethical guidelines of the Declaration of Helsinki. Informed written consent will be obtained from all subjects and their parents before study.

Inclusion criteria:

- children and adolescents (6-16 years);
- obese and normal-weight (control group) according to the Italian charts;
- both genders;
- diet naïve.

Exclusion criteria:

- heart and respiratory diseases;
- hypo/hyperthyroidism;
- GH deficiency:
- type 1 or type 2 diabetes;
- use of drugs (current or past): hormonal or interfering therapies (lipid-lowering, hypoglycemic, or antihypertensive treatment);
- renal or liver failure.

At the end of the study, obese subjects will be also divided into responders and non-responders to the treatment for the post-hoc analysis.

Clinical and anthropometric variables

All subjects will undergo a thorough medical history with special attention to the presence of cardiovascular risk factors and to the family history (I or II degree) of cardiovascular diseases and a clinical and anthropometric evaluation by trained physicians, using the Italian growth charts (25, *Appendix A*). Pubertal stages will be determined by using the criteria of Marshall and Tanner (26). Height will be measured to the nearest 0.1 cm by the Harpenden stadiometer and weight with light clothing to the 0.1 kg by using a manual scale. BMI will be calculated as body weight divided by squared height (kg/m2). BMI standard deviation score (BMISDS) will be calculated with the LMS method. Waist circumference will be measured with a soft tape, midway between the lowest rib margin and the iliac crest, in the standing position (27, *Appendix B*). Hip circumference will be measured over the widest part of the gluteal region, and the waist-to-hip ratio will be calculated. Systolic (SBP) and diastolic (DBP) blood pressure will be measured three times at the left arms by using a standard mercury sphygmomanometer and the mean value will be recorded and stratified according to paediatric percentiles of National High Blood Pressure Education Program Working Group on High Blood Pressure in Children and Adolescents (28, *Appendix C*).

Biochemical and metabolic evaluation

All subjects will be evaluated after 12 hours fasting for serum C-reactive protein (CRP), total- (Tc), HDL- (HDL-c) cholesterol, triglycerides (TG), AST, ALT, uric acid, creatinine, glucose, and insulin levels. Morning urine samples will be collected for the evaluation of microalbuminuria and urine creatinine. Plasma glucose levels (mg/dl; 1 mg/dl:0,05551 mMol/liter) will be measured by the gluco-oxidase colorimetric method (GLUCOFIX, by Menarini Diagnostici, Florence, Italy). Insulin (mUI/ml; 1 mUI/ml = 7.175 pmol/l) will be measured by chemiluminescent enzymelabelled immunometric assay (Diagnostic Products Corporation, Los Angeles, CA). Sensitivity: 2 mUI/ml. Intra- and inter-assay CV ranges: 2.5–8.3 and 4.4–8.6%. CRP (mg/dl), T-c (mg/dl; 1 mg/dl: 0.0259 mMol/l), HDL-c (mg/dl; 1 mg/dl: 0.0259 mMol/l), TG (mg/dl; 1 mg/dl: 0.0113 mMol/l), AST (U/l), ALT (U/l), uric acid (mg/dl; 1 mg/dl: 59.48 µMol/l), plasmatic creatinine (mg/dl; 1 mg/dl: 88 µMol/l) and microalbuminuria (mg/mmol urine creatinine; 1 mg/dL: 0.089 mMol/L) will be evaluated using standardized methods in the hospital's chemistry laboratory.

All obese and volunteer controls will perform an OGTT (1.75 g of glucose solution per kg, maximum 75 gr). Impaired fasting glucose (IFG) and impaired glucose tolerance (IGT) will be defined according to American Diabetes Association classifications as fasting plasma glucose of \geq 100 to 125 mg/dl, and as 2-h post-OGTT glucose of \geq 140 to 199 mg/dl, respectively. Also the

definition of diabetes will be performed according to the criteria of the American Diabetes Association (29). Blood samples during OGTT will be drawn for the determination of glucose and insulin every 30 minutes from 0 to 120 min. Insulin resistance will be estimated, in the basal state, by use of the homeostasis model assessment (HOMA-IR) =fasting glucose x fasting insulin/22.5. Insulin sensitivity will be estimated by the Matsuda $[10,000/\sqrt{3}]$ fasting glucose x fasting insulin) x (GmxIm)] and the quantitative insulin sensitivity check index (QUICKI), as defined by $1/\{\log[\text{fasting} \text{insulin } (\mu U/ml)] + \log[\text{fasting} \text{ glucose } (mg/dl)]\}$ (30). The stimulus for insulin secretion in the increment in plasma glucose as insulinogenic index (INS) will be calculated as the ratio of the changes in insulin and glucose concentration from 0 to 30 min. Beta-cell compensatory capacity was evaluated by the disposition index (DI) defined as the product of the Matsuda Index and INS (31).

Determination of interleukins, TNFα, plasminogen activator inhibitor-1, and adiponectin

IL-8, IL-10, IL-6, TNFα, plasminogen activator inhibitor-1 (PAI-1), and adiponectin will be measured using specific ELISA kit (R&D Systems) at the Laboratory of Clinical Pediatrics, Department of Health Sciences, University of Piemonte Orientale, Novara.

Plasma markers of oxidative stress measurement and plasmatic NO quantification

Blood samples will be taken for 3-nitrotyrosine, malondialdehyde (MDA), ROS generation, myeloperoxidase (MPO), GSH, and SOD measurement by specific kits. NO will be quantified from blood samples by using the Griess reagent. All these measures will be performed at the Laboratory of Physiology and Experimental Surgery, Department of Translational Medicine, University of Piemonte Orientale, Novara.

Cardiac function

A paediatrician, trained in echocardiography, will perform 2D-echocardiography. Transthoracic echocardiographic measurements will be performed with a GE Vivid 7 Pro, using a 3.0-mHz transducer for two-dimensional M-mode measurements of the left ventricular mass and a linear 3.0 to 11-mHz transducer for the vascular studies according to the recommendations of the American Society of Echocardiography (32). M-mode echocardiography will be used to measure left atrial and ventricular dimensions and left ventricular wall thickness, allowing for the calculation of left ventricular mass after correction for BSA; left ventricular mass and volumes will be calculated by Schiller's formula and Simpson's rule by two-dimensional echocardiography (33). Systolic function (ejection fraction [EF] and fractional shortening [FS]) and diastolic function (early (E) and late (A) diastolic peak flow velocities, and E/A ratio) will be measured using two-dimensional, M-mode, colour Doppler echocardiography. Interventricular septum (IVSt), left ventricular posterior wall thickness (LVPWt) and left ventricular end diastolic diameter (LVEDD) will be recorded, as well.

Aortic stiffness

The abdominal aortic diameter will be measured at maximum systolic expansion (Ds) and minimum diastolic expansion (Dd) by 2D-echocardiography. All aortic measurements will be made as previously described by Lacombe and Okubo (34,35). Aortic strain (S) will be calculated from the changes in aortic diameter using the formula $(S = (Ds-Dd)/Dd)$. Pressure strain elastic modulus (Ep) will be calculated from S, and the changes in brachial artery pressure will be determined by the formula (Ep=(Ps-Pd)/S). Pressure strain (Ep*), normalized by diastolic pressure, will be calculated using the formula ($Ep^* = Ep/Pd$). While S is the mean strain of the aortic wall, Ep and Ep^* are the mean stiffness of the aorta. Whereas S and Ep* are dimensionless ratios, Ep has the dimension newtons per square meter (i.e. force/unit area).

Carotid artery intima-media thickness (CIMT)

A vascular surgeon, blinded of the status of the patients will perform carotid sonography with the subject in supine position with a slight rotation of the neck using a linear 3.0- to 11-mHz transducer. The probe will be placed along the vessel axis to obtain a longitudinal scan of the common carotid arteries. CIMT will be measured 1.5 cm proximal to the carotid artery bifurcation, and the mean of three measurements of each artery will be reported. Data obtained will be compared with normative values, when possible (36).

Endothelium-dependent dilation of the brachial artery

Endothelium-dependent vasodilation will be measured in the right brachial artery in each subject by the same observer. After 10 min of rest, brachial artery blood flow and the diameter of the brachial artery 2 cm above the antecubital fossa will be determined by Doppler ultrasonography using a linear 3.0- to 11-mHz signal transducer. To induce hyperemia, a standard sphygmomanometer cuff will be applied to the forearm just proximal to the wrist joint and inflated to 300 mm Hg for 5 min. Blood flow and brachial artery diameter measurements will be repeated 45–60 sec after the release of sphygmomanometer pressure and will be calculated as the percentage change in brachial diameter and percentage change in flow.

Mitochondria morphology and function

Mitochondria will be isolated from monocytes. Ultrastructural analyses of mitochondria (through transmission electron microscope ZEISS EM 109) will be performed to assess morphologic mitochondrial changes (mitochondrial swelling, decrease in matrix density, possible difference in the sub-plasmalemmal and intrafibrillar sub-fraction of mitochondria, fission-fusion dynamic mitochondrial propriety, mitophagy etc). Moreover, mitochondria will be used for in vitro assays of mitochondrial oxygen consumption, complex I activity (NAD+/NADH), transmembrane potential and mitochondrial dynamic proteins expression (fusion and fission ratio through mitofusin 1 and 2 Western blot analysis). The evaluation mitochondria morphology and function will be performed at the Laboratory of Physiology and Experimental Surgery, Department of Translational Medicine, University of Piemonte Orientale, Novara.

Time course of measurements

All the evaluations previously described will be performed at baseline (T0) and after 6 (T1) and 12 months (T2) of an isocaloric Mediterranen balanced diet plus aerobic training.

Nutritional analysis and interventions

A well-trained and experienced clinical paediatric endocrinologist will assess food consumption in all subjects and will administer an isocaloric Mediterranen balanced diet in obese children. To assess food consumption, foods will be divided according to the classic basic food groups by the Italian Institute of Research on Food and Nutrition (38). Food frequencies questionnaires, validated for a wide range of ages (39), will be also completed by parents. The nutritional counselling will be performed at baseline and after 6 and 12 months, according to Italian LARN Guidelines for age and gender and the Italian food pyramid (38).

Moreover, obese subjects will undergo an exercise training regimen. Exercise will be conducted daily and will consist of 30 minutes of aerobic physical activity. Parents will record every day, on a specific questionnaire, the training performed.

Statistical analysis and power of the study

We will use an ITT analysis. The first main endpoint will be the difference of 10% of diastolic function between controls and obese. Estimates of the expected treatment effects will be obtained by using the SD of previous studies done in pediatric obesity (37). The second endpoint will be the variation of 20% of diastolic function in the obese group after 6 months of treatment. Estimates effects were obtained by using the SD of previous studies on probiotics in children (37). A sample

size of 50 individuals for each arm has been estimated to be sufficient to demonstrate those differences with a 90% power, a significance of 95%, and an estimated drop-out of 10% for the first outcome. A sample size of 125 obese individuals has been estimated to be sufficient to demonstrate those differences with a 90% power, a significance of 95%, and an estimated drop-out of 10% for the second outcome.

Data will be expressed as mean±SD, or percentages. The variation between groups will be evaluated by Student t-test. ANCOVA, correlation analysis and the stepwise regression model with two tailed probability values and 95% CI will be used when needed. P<0.05 will be considered significant. Statistical analyses will be performed with SPSS for Windows version 17.0.

References

- 1. Ebbeling CB, Pawlak DB. Childhood obesity: public-health crisis, common sense cure. *Lancet* 2002; 360:473- 482
- 2. Guo SS, Wu W, Chumlea WC, Roche AF. Predicting overweight and obesity in adulthood from body mass index values in childhood and adolescence. *Am J Clin Nutr* 2002; 76:653-658
- 3. Freedman DS, Khan LK, Dietz WH, Srinivasan SR, Berenson GS. Relationship of childhood obesity to coronary heart disease risk factors in adulthood: the Bogalusa Heart Study. *Pediatrics* 2001; 108:712-718
- 4. Young-Hyman D, Schlundt DG, Herman L, DeLuca F, Counts D. Evaluation of the insulin resistance syndrome in 5- to 10-year-old overweight/obese African-American children. *Diabetes Care* 2001; 24:1359- 1364
- 5. Strong JP, Malcom GT, McMahan CA, Tracy RE, Newman WP 3rd, Herderick EE, Cornhill JF. Prevalence and extent of atherosclerosis in adolescents and young adults: implications for prevention from the Pathobiological Determinants of Atherosclerosis in Youth Study. *JAMA* 1999; 281:727-735
- 6. Kortelainen ML. Adiposity, cardiac size and precursors of coronary atherosclerosis in 5- to 15-year-old children: a retrospective study of 210 violent deaths. *Int J Obesity* 1997; 21:691-697
- 7. McGill HC, McMahan A, Zieske AW, Tracy RE, Malcom GT, Herderick EE, Strong JP. Association of coronary heart disease risk factors with microscopic qualities of coronary atherosclerosis in youth. *Circulation* 2000; 102:374-379
- 8. Mehta SK, Richards N, Lorber R, Rosenthal GL. Abdominal obesity, waist circumference, body mass index, and echocardiographic measures in children and adolescents. *Congenit Heart Dis* 2009; 4:338-347
- 9. Dhuper S, Abdullah RA, Weichbrod L, Mahdi E, Cohen HW. Association of obesity and hypertension with left ventricular geometry and function in children and adolescents. *Obesity* 2011; 19:128-133
- 10. Chinali M, de Simone G, Roman MJ, Lee ET, Best LG, Howard BV, Devereux RB. Impact of obesity on cardiac geometry and function in a population of adolescents: the Strong Heart Study. *J Am Coll Cardiol* 2006; 47:2267-2273
- 11. Schuster I, Karpoff L, Perez-Martin A, Oudot C, Startun A, Rubini M, Obert P, Vinet A. Cardiac function during exercise in obese prepubertal boys: effect of degree of obesity. *Obesity* (Silver Spring) 2009; 17:1878- 1883
- 12. Sharpe JA, Naylor LH, Jones TW, Davis EA, O'Driscoll G, Ramsay JM, Green DJ. Impact of obesity on diastolic function in subjects<or=16 years of age. *Am J Cardiol* 2006; 98:691-693
- 13. Cote AT, Harris KC, Panagiotopoulos C, Sandor GGS, Devlin AM. Childhood Obesity and Cardiovascular Dysfunction. *J Am Coll Cardiol* 2013; 62:1309-1319
- 14. Meyer AA, Kundt G, Steiner M, Schuff-Werner P, Kienast W. Impaired flow-mediated vasodilation, carotid artery intima-media thickening, and elevated endothelial plasma markers in obese children: the impact of cardiovascular risk factors. *Pediatrics* 2006; 117:1560-1567
- 15. Kapiotis S, Holzer G, Schaller G, Haumer M, Widhalm H, Weghuber D, Jilma B, Röggla G, Wolzt M, Widhalm K, Wagner OF. A proinflammatory state is detectable in obese children and is accompanied by functional and morphological vascular changes. *Arterioscler Thromb Vasc Biol* 2006; 26:2541-2546
- 16. Beauloye V, Zech F, ThiMong HT, Clapuyt P, Maes M, Brichard SM. Determinants of early atherosclerosis in obese children and adolescents. *J Clin Endocrinol Metab* 2007; 92:3025-3032
- 17. Gilardini L, Pasqualinotto L, Di Matteo S, Caffetto K, Croci M, Girola A, Caffetto K, Croci M, Girola A, Invitti C. Factors associated with early atherosclerosis and arterial calcifications in young subjects with a benign phenotype of obesity. *Obesity* 2011; 19:1684-1689
- 18. Van Gaal LF, Mertens IL, Block CD. Mechanisms linking obesity with cardiovascular disease. *Nature* 2006; 444:875-880
- 19. Codoñer-Franch P, Valls-Bellés V, Arilla-Codoñer A, Alonso-Iglesias E. Oxidant mechanisms in childhood obesity: the link between inflammation and oxidative stress. Transl Res 2011; 158:369-384
- 20. Kershaw EE, Flier JS. Adipose tissue as an endocrine organ. *J Clin Endocrinol Metab* 2004; 89:2548-2556
- 21. Bełtowski J, Jamroz-Wiśniewska A, Widomska S. Adiponectin and its role in cardiovascular diseases. Cardiovasc Hematol Disord Drug Targets 2008; 8:7-46
- 22. Ouchi N, Walsh K. Adiponectin as an anti-inflammatory factor. *Clin Chim Acta* 2007; 380:24-30
- 23. Barseghian A, Gawande D, Bajaj M. Adiponectin and vulnerable atherosclerotic plaques. *J Am Coll Cardiol* 2011; 57:761-770
- 24. Grossini E, Prodam F, Walker GE, Sigaudo L, Farruggio S, Bellofatto K, Marotta P, Molinari C, Mary D, Bona G, Vacca G. Effect of monomeric adiponectin on cardiac function and perfusion in anesthetized pig. *J Endocrinol* 2014; 222:137-149
- 25. Cacciari E, Milani S, Balsamo A, Spada E, Bona G, Cavallo L, Cerutti F, Gargantini L, Greggio N, Tonini G, Cicognani A. Italian cross-sectional growth charts for height, weight and BMI (2 to 20 yr). *J Endocrinol Invest* 2006; 29:581-593
- 26. Tanner JM. Growth at adolescence. 2 edn. Oxford: Blackwell Scientific Publications 1961
- 27. McCarthy HD, Jarrett KV, Crawley HF. The developmental of waist circumference percentiles in British children aged 5,0 – 16,9 y. *Eur J Clin Nutr* 2001; 55:902-907
- 28. National High Blood Pressure Education Program Working Group on High Blood Pressure in Children and Adolescents. The fourth report on the diagnosis, evaluation and treatment of high blood pressure in children and adolescents. *Pediatrics* 2004; 114:555-576
- 29. American Diabetes Association. Diagnosis and classification of diabetes mellitus. *Diabetes Care* 2013; 36:67- 74
- 30. Matsuda M1, DeFronzo RA. Insulin sensitivity indices obtained from oral glucose tolerance testing: comparison with the euglycemic insulin clamp. *Diabetes Care* 1999; 22:1462-1470
- 31. Kahn SE, Prigeon RL, McCulloch DK, Boyko EJ, Bergman RN, Schwartz MW, Neifing JL, Ward WK, Beard JC, Palmer JP, et al. Quantification of the relationship between insulin sensitivity and beta-cell function in human subjects. Evidence for a hyperbolic function. *Diabetes* 1993; 42:1663-1672
- 32. Sahn DJ, DeMaria A, KIsslo J, Weyman A. Recommendations regarding quantification in M-mode echocardiography; results of a survey of echocardiographic measurements. *Circulation* 1978; 58:1072-1083
- 33. Schiller NB, Shaah PM, Crawford M, DeMaria A, Devereux R, Feigenbaum H, Gutgesell H, Reichek N, Sahn D, Schnittger I, et al. Recommendations for quantification of the LV by 2 dimensional echocardiography. *J Am Soc Echocardiogr* 1989; 2:358-367
- 34. Lacombe F, Dart A, Dewar E, Jennings G, Cameron J, Laufer E. Arterial elastic properties in man: a comparison of echo-Doppler indices of aortic stiffness. *Eur Heart J* 1992; 13:1040-1045
- 35. Okubo M, Ino T, Takahashi K, Kishiro M, Akimoto K, Yamashiro Y. Age dependency of stiffness of the abdominal aorta and the mechanical properties of the aorta in Kawasaki disease in children. *Pediatr Cardiol* 2001; 22:198-203
- 36. Jourdan C, Wuhl E, Litwin M, Fahr K, Trelewicz J, Jobs K, Schenk JP, Grenda R, Mehls O, Tröger J, Schaefer F. Normative values for intimamedia thickness and distensibility of large arteries in healthy adolescents. *J Hypertens* 2005; 23:1707-1715
- 37. Dhuper S, Abdullah RA, Weichbrod L, Mahdi E, Cohen HW. Association of obesity and hypertension with left ventricular geometry and function in children and adolescents. Obesity 2011; 19:128-133
- 38. Società Italiana di Nutrizione Umana. Livelli di assunzione raccomandati di energia e nutrienti per la popolazione italiana (LARN). Milan, Italy: S.I.N.U. 1996
- 39. Hammond J, Nelson M, Chinn S, Rona RJ. Validation of a food frequency questionnaire for assessing dietary intake in a study of coronary heart disease risk factors in children. *Eur J Clin Nutr* 1993; 47:242-250

Appendix A *Italian percentiles for weight and height (2-20 years). Girls. (Cacciari E. et al., 2006)*

Appendix A *Italian percentiles for weight and height (2-20 years). Boys. (Cacciari E. et al., 2006)*

Appendix A *Italian percentiles for BMI (2-20 years). Girls. (Cacciari E. et al., 2006)*

Appendix A *Italian percentiles for BMI (2-20 years). Boys. (Cacciari E. et al., 2006).*

Appendix B

Waist circumference, 90° percentile. (McCarthy HD. et al., 2001).

Appendix C

Blood pressure percentiles. Girls. (National High Blood Pressure Education Program Working Group on High Blood Pressure in Children and Adolescents, 2004).

Appendix C

Blood pressure percentiles. Boys. (National High Blood Pressure Education Program Working Group on High Blood Pressure in Children and Adolescents, 2004).

Current advances in the PhD program

Actually, 35 obese subjects who met the inclusion criteria, have been recruited at Division of Pediatrics, Department of Health Sciences, University of Piemonte Orientale, Novara. All these patients have been evaluated at baseline (T0).

In this population, 18 subjects were females (51.4%), 10 children were prepubertal (28,6%) and the mean age was of 11.3 ± 2.5 years.

All the children underwent clinical and anthropometric evaluation. The mean height was $150.8 \pm$ 2.5 cm, the mean weight was 69.4 ± 2.5 kg, the mean BMI was 29.8 ± 2.5 kg/m² and the mean BMI z-score was 2.25 ± 2.52 . 25 subjects (74.3%) showed a systolic or a diastolic blood pressure above the $90th$ percentile for age and sex and all the subjects (100%) showed a waist circumference above the $90th$ percentile for age and sex.

Biochemical and metabolic evaluation, included an OGTT, was performed in all the patients. 15 children (42.9%) showed an HDL-cholesterol below the 10th percentile and in 7 subjects (20%) triglycerides were above the $90th$ percentile for age and sex. Impaired fasting glucose (IFG) was detected in 2 children (5.7%) and impaired glucose tolerance (IGT) was found in 4 patients (11.4%). None of the children showed diabetes according to the criteria of the American Diabetes Association. 17 subjects (48.6%) met the International Diabetes Federation (IDF) criteria for metabolic syndrome and 13 patients (37.1%) met the criteria for metabolic syndrome proposed by the National Cholesterol Education Program (NCEP).

The day of the OGTT, at baseline, serum and plasma samples were collected for the determination of IL-8, IL-10, IL-6, TNFα, plasminogen activator inhibitor-1, and adiponectin*,* plasma markers of oxidative stress and plasmatic NO quantification, and isolation of mitochondria from monocytes.

All the 35 obese children underwent a transthoracic 2D-echocardiography. Left atrial and ventricular dimensions and left ventricular wall thickness have been recorded and left ventricular mass was calculated. The mean left ventricular mass index was 65.4 ± 15.2 g/m² and the mean left ventricular mass z-score was -0.05 ± 1.25 . Regarding systolic function, the mean ejection fraction (EF) was $69.3 \pm 8.6\%$ and the mean fractional shortening (FS) was $39.3 \pm 6.9\%$. Regarding diastolic function, the mean E/A ratio was 1.76 ± 0.54 . Interventricular septum (z-score, mean \pm SD: 0.10 ± 1.0), left ventricular posterior wall thickness (z-score, mean \pm SD: 0.45 ± 0.95) and left ventricular end diastolic diameter (z-score, mean \pm SD: -0.99 \pm 1.00) were recorded, as well.

A vascular echography was performed in all the subjects enrolled in the study. The abdominal aortic diameter was measured at maximum systolic expansion (Ds) and minimum diastolic expansion (Dd). The mean aortic strain was 0.29 ± 0.16 . Also pressure strain elastic modulus was (Ep) and pressure strain (Ep*) were calculated. Carotid artery intima-media thickness (CIMT) was measured in each children as well as endothelium-dependent dilation of the brachial artery, and femoral artery diameters.

Pubblications during PhD program

1. Cadario F, Savastio S, Magnani C, Cena T, Pagliardini V, Bellomo G, Bagnati M, Vidali M, Pozzi E, Pamparana S, Zaffaroni M, **Genoni G**, Bona G. High prevalence of vitamin d deficiency in native versus migrant mothers and newborns in the north of Italy: a call to act with a stronger prevention program. PLoS One, 2015; 10(6):e0129586, doi:10.1371/journal.pone.0129586. eCollection 2015

2. Monzani A, **Genoni G**, Cassone R, Binotti M, Ferrero F, Bona G. Hyperexcitability as the main sign of neonatal hypoxia. Minerva Pediatr, 2015; 67(3):276-277

3. De Rienzo F, Mellone S, Bellone S, Babu D, Fusco I, Prodam F, Petri A, Muniswamy R, De Luca F, Salerno M, Momigliano-Richardi P, Bona G, Giordano M; **Italian Study Group on Genetics of CPHD**. Frequency of genetic defects in combined pituitary hormone deficiency: a systematic review and analysis of a multicentre Italian cohort. Clin Endocrinol (Oxf), 2015; doi: 10.1111/cen.12849 [Epub ahead of print]. As a coauthor listed in the acknowledgements

Congress during PhD program

1. ESPE congress 2015 - **MODERATED POSTER SECTION** (October 2015) **Genoni G**, Esposito S, Agarla V, Monzani A, Castagno M, Raviolo S, Petri A, Prodam F, Bellone S, Bona G. "SHOX deficiency: clinical, radiological signs and value of screening scores"

2. Corso "Le cardiopatie congenite: dalla diagnosi alla profilassi respiratoria" (September 2015) – **Partecipant**

Courses and Seminars

- July 28, 2015. **Le cellule staminali nel danno renale acuto e nel trapianto di rene** (Prof Cantaluppi)
- July 9, 2015. **Miniworkshop on biotechnology for dermatology** (Prof Isidoro, Dr Rolin, Dr Lihoreau)